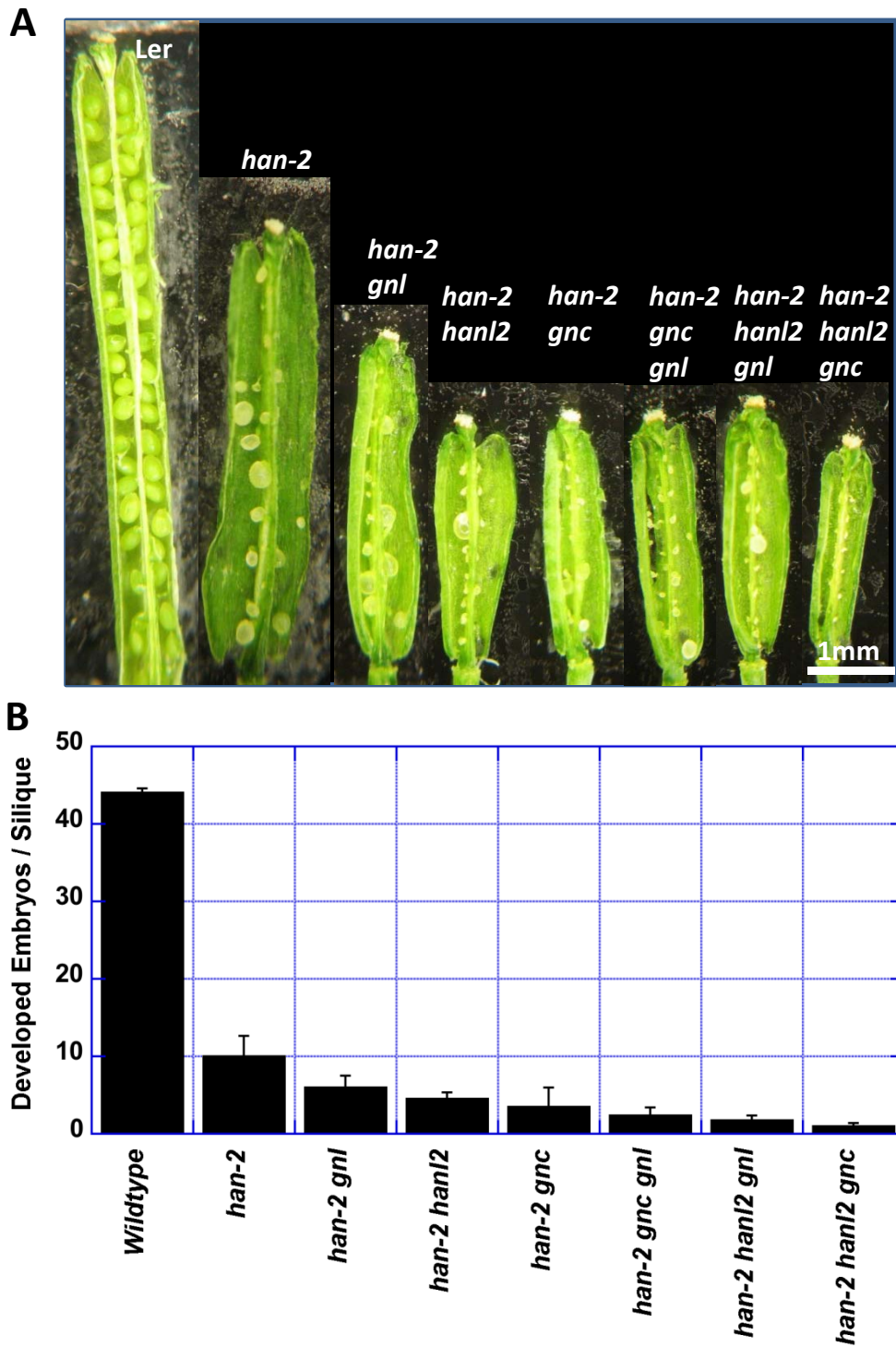
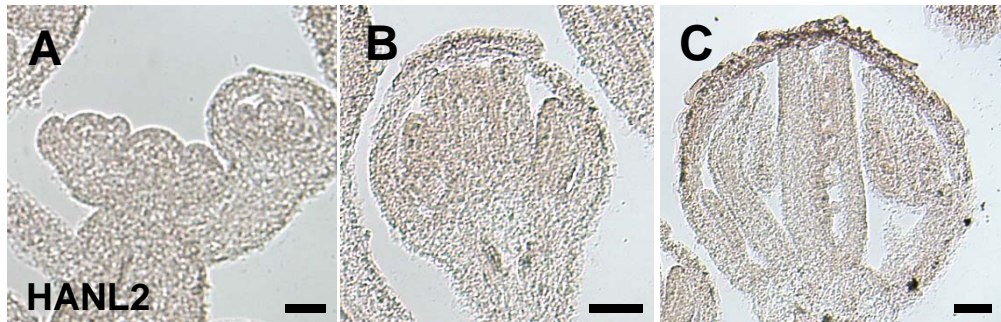


Supplemental Figure 1 . Gene ontology (GO) term enrichment ($P < 0.003$) in cluster 1 (A), cluster 2 (B), cluster 3 (C) and cluster 4 (D) of differentially expressed genes. GO terms are sorted based on p value.

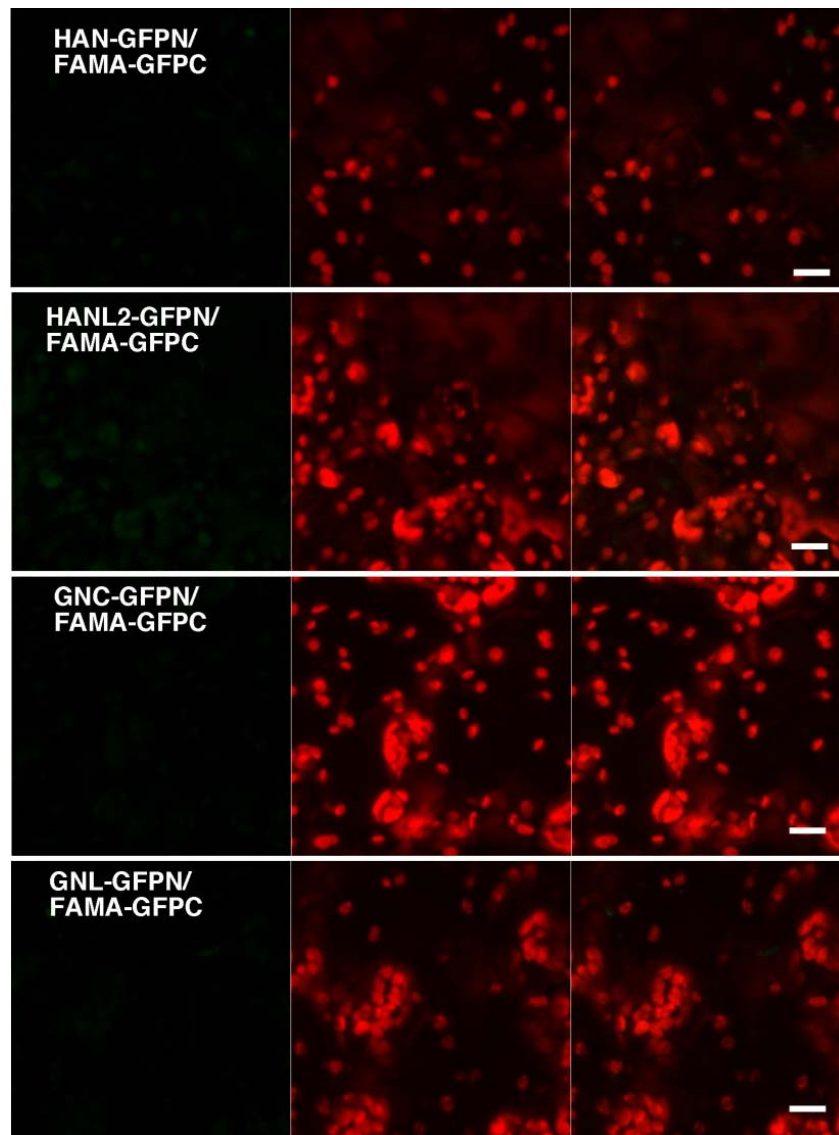


Supplemental Figure 2 . Seed yield is greatly reduced in the combinatory mutants of *HAN* and *GATA3* family genes.

(A) Opened siliques at similar developmental stages. (B) Average developed embryos per silique (n=10 siliques). Error bars represent standard deviation.



Supplemental Figure 3. *in situ* hybridization of *HANL2* in *hanl2* mutant flowers. A longitudinal section of the shoot apex (A), a stage 6 flower (B), and a stage 8 flower (C) reveals that *HANL2* signal is almost undetectable in the *hanl2* mutant allele SALK_138626. Bar = 50 μ m.



Supplemental Figure 4. Negative controls for interactions between HAN and GATA3 family proteins in BiFC experiments in transiently transformed *Nicotiana benthamiana* leaves. For each picture, a positive interaction is indicated by GFP fluorescence (green) in nuclei (left panel), the tobacco cells are seen via chlorophyll auto-fluorescence (red) (middle panel), and the merged channels are also shown (right panel). The label HAN-GFPN stands for the HAN fused with N-terminus half of GFP in frame, and similarly for the other constructs. Representative images of HAN-GFPN with FAMA-GFPC, HANL2-GFPN with FAMA-GFPC, GNC-GFPN with FAMA-GFPC and GNL-GFPN with FAMA-GFPC are shown, respectively. The pictures were taken under the same settings and each interaction was confirmed three times with independent infiltrations. Scale bar = 20 μ m.

Supplemental Table 1. Gene expression patterns of five non-GATA HAN targets in mutant plants and in the transient overexpression of HAN plants

Gene ID	Gene Name	DEX-4h			DEX-72h			DEX-9 days	<i>han-1</i>	<i>han-2</i>	<i>han-2 hanl2gnc</i>
		P-Value	Microarray	qPCR	P-Value	Microarray	qPCR	qPCR	qPCR	qPCR	qPCR
At4g27260	GH3.5	1.37E-08	3.60	2.14± 0.47	3.52E-01	1.70	7.20 ±0.12	2.82±0.20	-8.78± 0.80	-2.36±0.07	-1.18 ±0.02
At3g04730	IAA16	0.00E+00	-5.30	-2.34±0.16	3.88E-04	-2.00	-2.11 ±0.11	-1.21±0.11	-1.77±0.19	1.57±0.11	1.30±0.16
At3g16360	AHP4	1.53E-12	-5.00	-14.63±5.95	4.27E-02	-3.90	-175.1±39.9	-20.57±3.97	-0.60±0.05	-1.21±0.06	1.28 ±0.18
At4g23600	CORI3	1.78E-14	-2.70	-4.04±0.48	5.18E-34	-20.50	-93.64±35.4	-3.61± 0.82	-3.65±0.04	-1.53± 0.02	2.97±0.08
At3g15030	TCP4	7.92E-05	-1.80	-4.09±1.12	1.97E-07	-2.20	-5.90±1.07	-1.14± 0.26	-6.03±0.25	-2.63±0.11	-2.79±0.08

Supplemental Table 2. Primer information used in this study

Primers for qRT-PCR	
<i>HAN-F</i>	5'- TCTCCGCTAACAAGCCAAGT-3'
<i>HAN-R</i>	5'- GAGCCACGGAGTACCATTA -3'
<i>HANL2-F</i>	5'- CAACTTTGGATCATCAAGCAAG-3'
<i>HANL2-R</i>	5'- ACATCGTAAAATCGTGGACAA-3'
<i>GNC-F</i>	5'- GGTTGATGTCCCCAAAGATG -3'
<i>GNC-R</i>	5'- CACTTCGCCAAAGAGGAGTC -3'
<i>GNL-F</i>	5'- CCGCAGTCTCTGGCGTATC-3'
<i>GNL-R</i>	5'- GTGTTTACCTTTAGGGGCAAA -3'
<i>actin2-F</i>	5'- CCTTCGTCTTGATCTTGCGG-3'-3'
<i>actin2-R</i>	5'- AGCGATGGCTGGAACAGAAC -3'
<i>KNAT4-F</i>	5'- ACTGAGGAAGATAAGGCGAGGT-3'
<i>KNAT4-R</i>	5'- CGGTAGAAGAAGATGGATTGCT-3'
<i>RBE-F</i>	5'- AAGAAAGCCTAACGGAACCAA-3'
<i>RBE-R</i>	5'- CCAATGACCAACCCTAACATC-3'
<i>TCP4-F</i>	5'- ATCCGTTAGGGTTTGACACATC-3'
<i>TCP4-R</i>	5'- AGGAGTAGGAGGAGCGAACAG-3'
<i>MEE3-F</i>	5'- GTGCCATTCCCTGACTACAAG-3'
<i>MEE3-R</i>	5'- TGCTTCTCATCCTCTTTCTC-3'
<i>ATH1-F</i>	5'- TCCAAAACCTCCTTACCCTTA-3'
<i>ATH1-R</i>	5'- TGTTCAATTCGTCATACATCTC-3'
<i>EDA31-F</i>	5'- ATTTCAACTCCTCCTCGATTT-3'
<i>EDA31-R</i>	5'- TCTCATTCTCTCTCTCAGC-3'
<i>AFO-F</i>	5'- GTAAGTGTCCGATGTGGTTGC-3'
<i>AFO-R</i>	5'- CCTGGGGATTGAAGTAAGAGTG-3'
<i>KNAT2-F</i>	5'- TCGGAGAAAGCGACGTTGAT-3'
<i>KNAT2-R</i>	5'- TCGAATACGCGGAAGCAACTC-3'
<i>FUS3-F</i>	5'- GAAAATGTGGAAACCAAGGC-3'
<i>FUS3-R</i>	5'- AAGAGGCACAGACGCCGATAA-3'
<i>CYCD1:1-F</i>	5'- ACCTTTCTCGGGTTCTTTATCTC-3'
<i>CYCD1:1-R</i>	5'- ACTCGTTCGCTACACAGAGAATC -3'
<i>JAZ7-F</i>	5'- GCGAGTAGAGATGTGGAAGAGAA-3'
<i>JAZ7-R</i>	5'- CGGTGGTAAGGGGAAGTTGCT-3'
<i>SEP3-F</i>	5'- AGGGAGAGTAGAATTGAAGAGG-3'
<i>SEP3-R</i>	5'- ATTTGAGAAGATGATGAGAGCA-3'
<i>ATEXPA1-F</i>	5'- AGAGAGAGGGGCATGATAGTAATTT-3'
<i>ATEXPA1-R</i>	5'- TCAAGCACTCGAAGCACCACTT-3'
<i>ARR3-F</i>	5'- ACTCTACGGTGGAATCGCCTCT-3'
<i>ARR3-R</i>	5'- AAGCTAATCCGGGACTCCTCAT-3'
<i>ARR5-F</i>	5'- TTTTGCCTCCCGAGATGTTAG-3'
<i>ARR5-R</i>	5'- TACTATCATCAACAGCAAGAACATGA-3'
<i>ARR5-F</i>	5'- TTTTGCCTCCCGAGATGTTAG-3'
<i>ARR5-R</i>	5'- TACTATCATCAACAGCAAGAACATGA-3'
<i>ARR15-F</i>	5'- CCTAAGAGAATCCTACAAAACG-3'
<i>ARR15-R</i>	5'- CTTGGAAGATGGAGTGTCTGCA-3'
<i>RGL1-F</i>	5'- CGGATTCAAGAAAAGCCT-3'
<i>RGL1-R</i>	5'- ATTCTCGTCCTCTTGCTCC-3'

<i>GH3.5-F</i>	5'- TCGATCTGACGCTTGACCAAA-3'
<i>GH3.5-R</i>	5'- AAGATCTCCTCCAAGACTTGTCTTT-3'
<i>IAA16-F</i>	5'- GGTGGGAATCACGGAGGA-3'
<i>IAA16-R</i>	5'- GCTGGTGGTTTTACGACCTTCT-3'
<i>AHP4-F</i>	5'- CGGAAGGATGCTTGAGGACT-3'
<i>AHP4-R</i>	5'- TTGCCAACTGGAAATAATGT-3'
<i>COR13-F</i>	5'- GACTTTGATAGCGTCCGAGC-3'
<i>COR13-R</i>	5'- AAACCTCGTCAGAAACCACC-3'
<i>JAZ10-F</i>	5'- GCCGTCGTAGTTTCCGAGATA-3'
<i>JAZ10-R</i>	5'- CGAACGAGATTTAGCCGATGA-3'
<i>LEA14-F</i>	5'- AGGGAGATTGGAAGGGGAAG-3'
<i>LEA14-R</i>	5'- ATGGTTAGACCGATTGGAGC-3'
<i>IAA18-F</i>	5'- GAAATGACTCAACCACCTCCAAT-3'
<i>IAA18-R</i>	5'- ATCTTCTATGGATGATGGAAAGTCTC-3'
<i>IAA7-F</i>	5'- GGCTCCGTTGATCTCAAAAAC-3'
<i>IAA7-R</i>	5'- GCTGGCCTCCTCCGCAC-3'

Primers for *in situ* probes

<i>HAN-Sp6</i>	5'- GATTTAGGTGACACTATAGAATGCTCGTGAAACCTGCAAACCTGAA-3'
<i>HAN-T7</i>	5'- TGTAATACGACTCACTATAGGGACCGGACTTGGCTTGTTAG-3'
<i>HANL2-SP6</i>	5'- GATTTAGGTGACACTATAGAATGCTCACAACACACACTTCTTTCTCC-3'
<i>HANL2-T7</i>	5'- TGTAATACGACTCACTATAGGGGAAGTGGTGTGCGAGTTGG-3'
<i>GNC-SP6</i>	5'- GATTTAGGTGACACTATAGAATGCTTTGGACTCTTTTGCCGTCTAT-3'
<i>GNC-T7</i>	5'- TGTAATACGACTCACTATAGGGATGGTGATCCTCGTCGAAATTA-3'
<i>GNL-SP6</i>	5'- GATTTAGGTGACACTATAGAATGCTTTCGCTTCTTTGGATCCTC-3'
<i>GNL-T7</i>	5'- TGTAATACGACTCACTATAGGGCGGAGCAAATCCTAATCACG-3'

Primers for *BiFC*

<i>HANL2c/5CACC</i>	5'-CACCATGGGTTTCTCAATGTTCTTCT-3'
<i>HANL2c/3stop</i>	5'-TCACATCGTAAATCGTGGACA-3'
<i>GNCc/5CACC</i>	5'-CACCATGGATTCAAATTTTCATTACT-3'
<i>GNCc/3stop</i>	5'-TCAACCGTGAACCATTCATAC-3'
<i>GNLc/5CACC</i>	5'-CACCATGGGTTCCAATTTTCATTACA-3'
<i>GNLc/3stop</i>	5'-TCACCCGTGAACCATTCGGTGC-3'
<i>HANL2c/5</i>	5'-ATGGGTTTCTCAATGTTCTTCT-3'
<i>HANL2c/3</i>	5'-CATCGTAAATCGTGGACA-3'
<i>GNCc/5</i>	5'-ATGGATTCAAATTTTCATTACT-3'
<i>GNCc/3</i>	5'-ACCGTGAACCATTCATAC-3'
<i>GNLc/5</i>	5'-ATGGGTTCCAATTTTCATTACA-3'
<i>GNLc/3</i>	5'-CCCGTGAACCATTCGGTGC-3'
<i>HANc/5CACC</i>	5'-CACCATGATGCAGACTCCGTACACTAC-3'
<i>HANc/3stop</i>	5'-TCATCTGGTAAAGTCATGGACAAGA-3'
<i>HANc/5</i>	5'-ATGATGCAGACTCCGTACACTAC-3'
<i>HANc/3</i>	5'-TCTGGTAAAGTCATGGACAAGA-3'

Primers for ChIP-PCR

<i>HANp1-F</i>	5' - AACATCCGATGAAATTGGAA-3'
<i>HANp1-R</i>	5' - AGATGACGGTTGTCCAAAAA-3'
<i>HANp2-F</i>	5' - ATCGAGAGTCCATGTTTTCAATAAA-3'
<i>HANp2-R</i>	5' - CTCAGCCTCATGTGGTTAATGAT-3'
<i>HANp3-F</i>	5' - TTATTTTCTGTAAATTTCTCTCA-3'
<i>HANp3-R</i>	5' - CATTGCTAGCTTCCAAATATCA-3'

<i>HANp4-F</i>	5' - ACTAGCAAACCTTCGCTTT-3'
<i>HANp4-R</i>	5' - TGTCAGTGTGTGATTGTTTTGA-3'
<i>HANp5-F</i>	5' - TGGCTGCCTAAAGAACTTG-3'
<i>HANp5-R</i>	5' - GAATGGAGAAGAAGGTCTACA-3'
<i>HANp6-F</i>	5' - CGTCCCCACATATTCTATTGATTT-3'
<i>HANp6-R</i>	5' - TCTACTGGTTTGAAAACCTAAGTG-3'
<i>UBQ10-F</i>	5'- TCCAGGACAAGGAGGTATTCCTCCG-3'
<i>UBQ10-R</i>	5'- CCACCAAAGTTTTACATGAAACGAA-3'
<i>GNCp1-F</i>	5'- TTGTCTATGAAGTTTCTATCGTCTAT-3'
<i>GNCp1-R</i>	5'- GCTTTTATATGGGATCAGATTATG-3'
<i>GNCp2-F</i>	5'- TGGCTTATCACAACAAAGTCTATAAT-3'
<i>GNCp2-R</i>	5'- AAAATGAATATCTAACTAGCGTCTG-3'
<i>GNCp3-F</i>	5'- GACGGCAAAAAGAGTCCAAG-3'
<i>GNCp3-R</i>	5'- TGAGTCCATGGCACCCATA-3'
<i>GNCp4-F</i>	5'- CCGGACTTGTCCGAAAGTAG-3'
<i>GNCp4-R</i>	5'- GACGGCAAAAAGAGTCCAAG-3'
